Adjuvant effects of *Lactobacillus casei* added to a renutrition diet in a malnourished mouse model

**PAOLA GAUFFIN CANO**¹, **GRACIELA AGÜERO**³ AND **GABRIELA PERDIGON**¹,²

². Cátedra de Inmunología, Instituto de Microbiología. Facultad de Bioquímica, Química y Farmacia. Universidad Nacional de Tucumán.
³. Cátedra de Análisis Clínico, Instituto de Bioquímica Aplicada. Facultad de Bioquímica, Química y Farmacia. Universidad Nacional de Tucumán.

**Key words:** malnutrition, *Lactobacillus casei*, mucosal immunity, gut barrier, renutrition diet.

**ABSTRACT:** Nutritional deficiencies are associated with impaired immune response, affecting the body’s defence mechanisms. It is also known that Lactic Acid Bacteria (LAB) and fermented products such as yogurt have immunopotentiator activity and nutritional properties, and could thus be used as a valuable supplement in a renutrition diet. The aim of this study was to determine, in a non-severe malnutrition model, the effective dose of *Lactobacillus casei* (*L.casei*), which when is used as an adjuvant in a renutrition diet, would modulate the mucosal immune system and induce recovery of the integrity of the intestinal barrier. The experiments were performed on groups of malnourished and renourished BALB/c mice. They received after milk renutrition a supplement of different doses and periods of *L. casei* feeding. We measured body weight; hematologic values and serum proteins. We also characterized small intestine immunoglobulin secreting cells, intraepithelial leukocytes, mastocytes and goblet cells. Structural and ultrastructural studies were performed. Our results suggest that impaired gut barrier and mucosal immune function produced by malnutrition can be reversed by *L. casei* and that the dose of 10⁷ cfu/day/mouse administered during 5 consecutive days was the optimal one for recovery of the gut mucosal immune system. The clinical significance of these findings suggests ways for improving mucosal immunity, and generating protection against enteropathogens in hosts immunosuppressed by malnutrition.

**Introduction**

The gastrointestinal tract is undoubtedly the most exposed body site with respect to pathogenic microorganisms and non-viable materials, including antigens and carcinogens. As the major immunological organ in the body, it contains cells that absorb, process and present antigen to B and T cells of various types, and antibody producing cells. The intestinal mucosa provides both a physiologic and immunologic barrier to a wide range of microorganisms and foreign substances, by means of extensively interacting populations of cells and their products which are involved in innate and acquired defences of the body’s largest surface area (Deitch, 1992; Karlsson *et al.*, 1996; Brandtzaeg, 1998; Roux *et al.*, 2000). In general, the mucosal immune system is homeostatic, but when an imbalance occurs in the regulation of this response, gut barrier dysfunction and inflammatory bowel disease are observed.
The relationship between nutrition, development and immunity has been the focus of increasing research in the last two decades. Thus, the function of many cells in the immune system relies on metabolic pathways that in turn depend upon various nutrients as critical cofactors, which in turn affects body defense mechanisms (MacDermott, 1993).

It is now established that nutritional deficiency is commonly associated with an impaired immune response, particularly cell-mediated immunity, phagocyte function, cytokine production, secretory antibody response, antibody affinity, and the complement system (Chandra and Wadhwa, 1993; Chandra, 1997). Non-specific mechanisms that include intestinal flora, anatomical barriers (mucosa and epithelium), secretory substances such as lysozymes and mucus are affected by malnutrition (Sherman et al., 1985). In fact, malnutrition is the commonest cause of immunodeficiency in underdeveloped countries. It is suggested that impaired systemic and mucosal immunity contribute to the increased frequency and severity of intestinal infection seen in undernourished individuals.

In protein-energy malnutrition (PEM), most of the host defence mechanisms are breached depending on the severity of protein deficiency relative to energy, allowing microbes to invade and produce clinical infection, which is more severe and prolonged (Chandra, 1996; Scrimshaw and Sangiovanni, 1997).

That the detrimental effects of severe malnutrition on the mucosal immune system can be reversed with adequate refeeding is of further clinical importance. Similarity, refeeding rapidly restores the morphology and function of intestine resulting in repair of gut atrophy and normalization of intestinal permeability (Castillo et al., 1991; Poulain et al., 1991; Boza et al., 1999). Milk and milk products represent important sources of dietary proteins for humans, and these proteins are known to serve as a source of biologically active peptides. It was well demonstrated that the presence of a number of growth factors and hormones in the milk of various species including human and bovine, have beneficial effects on the host and so it suggests its potential use in the renutrition process (Koldovsky, 1989; Meisel and Bockelmann, 1999; Matar et al., 2000).

The gut microflora is an important constituent of the intestinal mucosa barrier and this has led to the concept of probiotic therapy, i.e. the application of potentially beneficial microorganisms (Fuller, 1992). LAB with probiotic activity were used in various fermented foods since antiquity. Probiotic is a microbial dietary adjuvant that beneficially affects the host physiology by modulating mucosal and systemic immunity, as well as improving nutritional and microbial balance in the intestine tract. The probiotic spectrum of activity can be divided into nutritional, physiological, and antimicrobial effects. LAB are also potential adjuvants, and their oral administration triggers mucosal and systemic immune responses. Various effects ascribed to LAB are summarized as follows: improvement of the nutritional quality of food and feed; stimulation of vitamin synthesis and enzyme production; stabilization of gut microflora and competitive exclusion of enteric pathogens; enhancement of innate host defences by production of antimicrobial substances; reduction of serum cholesterol by assimilation mechanisms; decreased risk of colon cancer by detoxification of carcinogens and tumor suppression by modulation of cell-mediated immunity (Gerritse et al., 1990; Perdigón et al., 1990; 1994; Naidu et al., 1999). Probiotics can be used as innovative tools for treating dysfunctions of the gut mucosal barrier including acute gastroenteritis, food allergy, and inflammatory bowel disease.

In our laboratory it was demonstrated that L. casei was able to induce a secretory immune response, which was related to the dose administered to healthy mice (Perdigón et al., 1991). However when we used an experimental model of severe malnutrition, we showed that yogurt and L. casei were not able to increase the protective intestinal mechanisms to the same levels found in the well-nourished host (Perdigón et al., 1995; Agüero et al., 1996; Perdigón and Oliver, 2000). Therefore, in order to avoid harmful effect at intestinal level by an overstimulation of the immune system we recommended the use of LAB in the malnourished mice after mucosal recovery by adequate refeeding.

The aim of this study was to determine, in a non-severe malnutrition experimental model, the effective dose of L. casei used as adjuvant in a renutrition diet required to modulate the mucosal immune system and to repair the integrity of intestinal barrier.

**Material and Methods**

**Diets**

The protein-free diet was supplemented in order to fulfill nutritional requirements. It was composed and designed as follows: vitamin mixture (2.2%) (ICN, Biomedicals, Inc-1263 South Chillicothe Road, Aurora, Ohio 44202), salt mixture (4%) (ICN, Biomedicals, Inc-1263 South Chillicothe Road, Aurora, Ohio 44202), and corn oil (5%).
**Animals**

Weaning BALB/c mice obtained from closed colony of the breeding unit kept at CERELA were housed in cages at 23°C with a 12 h light/dark periods. They had free access to protein-free diet supplemented with vitamins, minerals and essentials fat free acid for 21 days. At the end of this period, the animals that had lost about 35-55% of their weight compared with the well-nourished controls were selected for the experiments. The mice were split into two groups. One group was the malnourished (M-N) and the other was refeeding with 10%-non-fat-milk as the source of protein (NFM) for 7 days; this was the re nourished group (Re7d). Comparative assays between M-N and Re7d animals were affected under identical conditions.

Experiments were performed with 10 animals for weight body determinations and 5 animals for the other assays.

**Microorganism feeding procedure**

The bacterial strain used for experiments was *Lactobacillus casei* var *casei* CRL 431 from CERELA culture collection. It was cultured for 8 h at 37°C in MRS (De Man et al., 1960) broth (Oxoid Ltd., U.S.A.) and harvested by centrifugation at 5,000 rpm for 10 min, and washed three times with sterile saline solution. Mice were daily fed with culture of LAB at a concentration of 10⁶, 10⁷ and 10⁸ colony forming units (cfu)/day/animal for 2, 5 or 7 consecutive days. The cultures were administered at 20% v/v in the drinking water to M-N groups and in NFM 10% to Re7d group.

We used three control groups: well nourished which received conventional diet (W-N), M-N (21 days protein-free diet) and Re7d controls (mal-nourished mice reno nourished for 7 days with 10% NFM).

**Body weight determinations**

It was determined at the end of each treatment period in groups of 10 mice and expressed as grams

**Hematological and serum total protein determinations**

They were performed in all experimental and control groups. Peripheral blood was recovered by cardiac puncture. The hematocrit (HTO) and number of leukocytes were determined by the hemacitometric method. PMN and lymphocytes populations were differentiated on smears stained with Giemsa solution. The total protein concentration was determined using Bradford technique (1976).

**Tissue sections**

The small intestine was removed at the end of each treatment period and processed by modified Saint-Maries’s technique (1962). Briefly, tissues were fixed in 95% ethanol for 24 hours at 4°C, and then were de-

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**TABLE 1.**

Effect of different doses and periods of *Lactobacillus casei* feeding on body weight of malnourished and reno nourished mice.

<table>
<thead>
<tr>
<th>Body Weight (g) ± SD</th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>2</td>
<td>5</td>
<td>7</td>
</tr>
<tr>
<td><strong>Test groups</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lactobacillus casei</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(cfu/day/mouse)</td>
<td>10⁶</td>
<td>10.1±1.3</td>
<td>10.7±0.9</td>
</tr>
<tr>
<td></td>
<td>10⁷</td>
<td>10.4±13</td>
<td>13.0±1.5</td>
</tr>
<tr>
<td></td>
<td>10⁸</td>
<td>9.8±1.1</td>
<td>11.9±0.2</td>
</tr>
</tbody>
</table>
| **Values for well-nourished groups**= 25.8±1.5 g; M-N control groups= 10.4±1.2** g and Re7d control groups= 12.1±1.5** g

M-N: malnourished; Re7d: reno nourished.

¹Values are means ± SD of 10 mice per group. ²Significantly different from Re7d control. ³Significantly different of well-nourished control. *P<0.05. **P<0.01 (ANOVA and Student’s Test)
hydrated in 3 changes of absolute alcohol and cleared by passing through 3 consecutive baths of xylene at 4°C for 45 min each one. The tissue was embedded in paraffin at 56°C for 3 to 6 h. Sectioning was carried out as usual, and tissue sections (3-4 µm) were placed on glass slides.

**Number of IgA, IgG and IgM secreting cell determinations**

The number of IgA, IgG and IgM secreting B cells were determined on histological slices from small intestine by direct immunofluorescence technique. It was performed using the respective monospecific antibodies (α, γ and µ-chain specific) conjugated with fluorescein isothiocyanate (FITC) (Sigma, Saint Louis, Missouri 63103, USA). Histological samples were incubated with 0.1 ml of different antibodies at 1/60 dilutions for IgA, and 1/100 for both IgG and IgM for 30 min at 37°C in humidified chamber. Then they were washed three times with 0.01M-phosphate-buffered saline (PBS), pH 7.2. The slices were mounted in glycerol:PBS. The number of positive fluorescent cells was expressed as number of secreting cells per 10 fields (magnification 100X).

**TABLE 2.**

Effect of different doses and periods of *Lactobacillus casei* feeding on the hematological values of malnourished and renourished mice.

<table>
<thead>
<tr>
<th>Test and control groups</th>
<th>Hematocrit (%)</th>
<th>White cells/µl</th>
<th>PMN (%)</th>
<th>Lymphocytes (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Hematocrit</td>
<td>White cells</td>
<td></td>
<td></td>
</tr>
<tr>
<td>L. casei</td>
<td>M-N</td>
<td>Re7d</td>
<td>M-N</td>
<td>Re7d</td>
</tr>
<tr>
<td>Days of feeding</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>10⁶ (cfu/day/mouse)</td>
<td>41±0.9</td>
<td>45±1.3</td>
<td>3500±106²</td>
<td>3300±100</td>
</tr>
<tr>
<td>5</td>
<td>38±0.8</td>
<td>40±0.8</td>
<td>3900±100²</td>
<td>3900±89²</td>
</tr>
<tr>
<td>7</td>
<td>37±0.9</td>
<td>42±1.1</td>
<td>3900±100²</td>
<td>3600±90³</td>
</tr>
<tr>
<td>10⁷ (cfu/day/mouse)</td>
<td>40±1.2</td>
<td>39±1.5</td>
<td>3390±107</td>
<td>3400±100</td>
</tr>
<tr>
<td>2</td>
<td>37±0.9</td>
<td>42±1.0</td>
<td>3100±80²</td>
<td>3800±120³</td>
</tr>
<tr>
<td>10⁸ (cfu/day/mouse)</td>
<td>43±1.0</td>
<td>42±1.0</td>
<td>3550±103²</td>
<td>3150±110³</td>
</tr>
<tr>
<td>5</td>
<td>40±0.9</td>
<td>43±1.7</td>
<td>37800±90²</td>
<td>3600±100³</td>
</tr>
<tr>
<td>7</td>
<td>39±1.1</td>
<td>45±1.0</td>
<td>38500±100</td>
<td>4000±100³</td>
</tr>
<tr>
<td>M-N and Re7d controls</td>
<td>42±1.0⁴</td>
<td>39±0.9⁴</td>
<td>2800±106⁴</td>
<td>3200±106⁴</td>
</tr>
<tr>
<td>W-N control</td>
<td>55±10</td>
<td>4800±100</td>
<td>15±1</td>
<td>82±0.9³</td>
</tr>
</tbody>
</table>


1Values are means of n = 5 per group ± SD. 2Significantly different from M-N control (P<0.01). 3Significantly different from Re7d control (P<0.01). 4Significant differences with W-N control (P<0.01). 5Significant differences with M-N control (P<0.01). 6Significant differences with Re7d control. *P<0.05, **P<0.01 (ANOVA and Student’s Test).

**FIGURE 1.** Number of IgA, IgG and IgM secreting cells present in the small intestine of malnourished and renourished mice treated with different doses and periods of *Lactobacillus casei* feeding, by immunofluorescent test. The values are means ± SD of two small intestine’s sections of each of 5 experimental and control mice.

W-N: well-nourished control. M-N: malnourished control. Re7d: renourished control. 1Significant differences with W-N control (P<0.01). 2Significant differences with M-N control (P<0.01). 3Significant differences with Re7d control. *P<0.05, **P<0.01 (ANOVA and Student’s Test).

●: IgA · cells; ■: IgG · cells; ▼: IgM · cells.
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Malnourished

Renourished

10^6 cfu/day/mouse

10^7 cfu/day/mouse

10^8 cfu/day/mouse

Feeding period

Feeding period

Number secreting cells/10 fields

W-N, M-N, 2d, 5d, 7d

W-N, M-N, Re7d, 2d, 5d, 7d

**p < 0.01

* p < 0.05
Number of intraepithelial leukocytes (IEL), mast cells, and goblet cells

Histological slices were stained with Hematoxilin-eosin for IEL. For mastocytes and goblet cells slices were stained with 1% Alcian Blue 8GX (Merck, F.R. Germany, Darmstadt D-6100) in 3% acetic acid - 0.5% Safranin O (Sigma, Saint Louis, Missouri 63103, USA) in 0.01M HCl. This was performed as described by Koretou (1988). The number of cells was expressed per 10 fields (magnification 100X).

Preparation of samples for electronic microscopy for ultrastructural studies

At the end of each treatment period, the mice were sacrificed by cervical dislocation. Peyer’s patches and small intestine were carefully removed. Tissues were fixed in 40% formaldehyde and 10% glutaraldehyde in phosphate buffer, pH 7.2. Specimens were then washed in sodium phosphate buffer and fixed in 10% OsO4, dehydrated in ethanol, cleared in propylene oxide and finally embedded in low-viscosity medium. Thin sections were stained with saturated uranyl acetate in 50% ethanol and 4% citrate. Sections were examined by transmission electron microscopy and the micrographs were produced at 4,900 or 12,800 magnification. Specimens of control mice were processed in the same way.

Statistical analysis

Data were summarized using descriptive statistics such as the mean and standard deviations by Student’s test. Statistical comparisons of the treatment versus control groups were analyzed by ANOVA test.

Results

Body weight

The body weight of mice from the M-N and Re7d control groups was significantly lower (P<0.01) than that of mice from the well-nourished group. The oral supplementation with different doses of L. casei to M-N groups did not produce a significant difference with respect to M-N control. When the effect of L. casei treatment on the body weight of Re7d group was examined, all the doses studied produced significant increases at 7 days of feeding, and this was also observed on the 5th day of supplementation of the diet with 10⁷ cfu/day/mouse compared with Re7d control group (Table 1).

Number of IgA, IgG and IgM secreting cells

As shown in Figure 1 the malnutrition produced a remarkable decrease in the number of IgA secreting cells associated with the lamina propria of the small intestine, and the renutrition diet enhanced the IgA+ cells near to the values of well-nourished control mice. When we analyzed the effect of oral supplementation of diet with different doses of L. casei to M-N groups, L. casei induced a significant (P<0.01) increase in the number of IgA+ cells compared with M-N control at 7 days of treatment for 10⁶ and 10⁷ cfu/day/mouse, and 5 days of administration of 10⁶ cfu/day/mouse when compared to the M-N control. The values of IgA+ cells obtained by
TABLE 3.

Effect of different doses and periods of *Lactobacillus casei* feeding on the serum total protein concentration of malnourished and renourished mice.

<table>
<thead>
<tr>
<th>Test and control groups</th>
<th>Total serum protein (g/l)(^1)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Lactobacillus casei</em></td>
<td>Days of feeding</td>
</tr>
<tr>
<td>10⁶ (cfu/day/mouse)</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td>5</td>
</tr>
<tr>
<td></td>
<td>7</td>
</tr>
<tr>
<td>10⁷ (cfu/day/mouse)</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td>5</td>
</tr>
<tr>
<td></td>
<td>7</td>
</tr>
<tr>
<td>10⁸ (cfu/day/mouse)</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td>5</td>
</tr>
<tr>
<td></td>
<td>7</td>
</tr>
<tr>
<td>M-N and Re7d controls</td>
<td>40±8(^2)</td>
</tr>
<tr>
<td>Well-nourished control group</td>
<td>61±4</td>
</tr>
</tbody>
</table>

\(^1\)Values are means ± SD of determinations of 5 mice per group. \(^2\)Significantly different from W-N control (P<0.01). M-N: malnourished. Re7d: renourished. No significant variations was observed between treated malnourished and renourished mice compared with their respective control.

renutrition did not suffer significant modifications with the oral administration of *L. casei* except for the dose of 10⁶ cfu/day/mouse during 5 days. This dose also gave a slight increase in the number of IgG secreting cells, which was significant (P<0.05) at 7 days of treatment compared with Re7d control mice. The effect of *L. casei* administration on IgM secreting cells in both malnourished and renourished groups was not significant. Moreover, the number of these cells remained lower than those from well-nourished control mice. These results are expressed in Figure 1.

*Number of intraepithelial leukocytes (IEL), mast cells, and goblet cells*

The malnutrition significantly decreased the number of IEL and the renutrition diet significantly enhanced (P<0.05) those values compared with the well-nourished control. The feeding with *L. casei* induced a significant decrease of this parameter in almost all the doses used with respect to M-N and Re7d control (P<0.05, P<0.01, respectively). When we analyzed the number of goblet cells we observed that the diminished values obtained by the malnutrition diet were slightly increased by the oral administration of *L. casei* compared with M-N control. However this rise was significant only at 5 and 7 days of feeding with 10⁷ cfu/day/mouse, and dose of 10⁶ cfu/day/mouse during 7 days. The renutrition diet enhanced the number of these cells near to the well-nourished mice, and the different doses of *L. casei* produced a slight decrease at concentrations of 10⁶ and 10⁷ cfu/day/mouse. The significant decrease (P<0.01) seen in mast cells in M-N and Re7d mice compared with well-nourished control was not significantly affected by *L. casei* feeding.

*Histological and ultrastructural studies of the small intestine*

After malnutrition diet, an important alteration of size and number of villi per unit area was observed in hematoxilin-eosin stained slices of small intestine examined by light microscopy. When the mice were renourished, a marked recovery in the intestinal epithe-
lum was seen and the supplementation with *L. casei* at 10⁷ cfu/day/mouse during 5 days showed fewer signs of inflammation and edema in the villi than other doses and periods of treatment in relation to M-N and Re7d control, respectively (Fig. 4 a, b, c, d). By transmission electron microscopy we studied the effect of malnutrition on intestinal cells. There was an increase in the number of microvilli per cell over the well-nourished control but were narrower and longer than those of the microvilli in renourished control and treated mice. Animals whose diet was supplemented with dose of 10⁷ cfu/day/mouse showed microvilli which were more uniform with prominent rugose endoplasmatic reticulum in enterocytes (Fig. 3 a, b, c, d).

**Discussion**

The primary role of the gastrointestinal tract is to digest and absorb nutrients according to the metabolic requirements and demands for normal host growth and development. In addition, the intestinal mucosa provides a protective host defence against the constant presence of antigens from food and microorganisms in the gut lumen. The loss of nutrients that are essential for epithelial integrity, may affect the function of the gastrointestinal mucosa (Kensch, 1990; Lunn and Northrap-Claves, 1991; Welsh et al., 1999). The epithelial permeability for luminal antigens affected by malnutrition is probably an important primary or secondary event in the pathogenesis of several mucosal diseases, including adverse immune reactions to foods (Brandtzaeg, 1996).

The nutritional manipulation of the immune system is now thoroughly studied and it brings the promise of using diet and nutrition as innovative powerful tools to reduce illness and death caused by infection.

In the present research we demonstrated positive biological effects in the immunosuppressed model by malnutrition when we used *L. casei* as an adjuvant added to a renutrition diet. A similar strategy has previously been suggested by German et al. (1999) for the functional foods.

The use of such bacteria as probiotic supplements needs a knowledge of the effective doses to avoid an overstimulation of the gut mucosa immune system by the higher entry of antigens. In the development of this study we used three daily different doses of *L. casei* feeding: 10⁶, 10⁷ and 10⁸ cfu/day/mouse during 2, 5 or 7 consecutive days of administration. We demonstrated that the concentration of 10⁷ cfu/day/mouse during 5 days was the optimal dose to improve the intestinal barrier and the mucosal immune system in our malnutrition model. This choice was based on counts of the number of IgA+ cells (Fig. 1) and histological considerations from hematoxilin-eosin and transmission electron microscopy studies (Figs. 3, 4).

After entering the intestine, live and biologically active LAB particles may generate specific or non-specific immune responses of gut-associated lymphoid tissue (GALT) (Vintini et al., 2000). These bacteria can penetrate the gut wall either by translocation through the epithelial barrier or by encountering the Peyers’ patches (Perdigón et al., 2000). Subsequently, they reach the mesenteric lymph nodes and spleen, which stimulates immunocompetent cells to secrete immunoglobulins. The main function of the secretory antibody system is, in cooperation with innate mucosal mechanisms, to inhibit colonization and invasion of pathogens (Goldblum et al., 1996; Mestecky et al., 1999). In addition, induction of S-IgA responses has been shown to induce significant resistance to infection (Onorato et al., 1991). Therefore, it may be hypothesized that IgA antibodies contribute to local homeostasis not only by performing immune exclusion on the mucosal surface but also by trapping soluble antigens in the lamina propria (Brandtzaeg, 1998). Functional advantages of S-IgA over other immunoglobulin isotypes include its multi-valency (4-8 antigen binding sites) (Renegar et al., 1998). In addition to a specific antibody dependent

**FIGURE 2.** Number of intraepithelial leukocytes (IEL), and goblet and mast cells present in the small intestine of malnourished and renourished mice with different dose and periods of *Lactobacillus casei* feeding by hematoxilin-eosin and Alcian blue staining respectively. The values are means ± SD of two small intestine’s sections of each of 5 experimental and control mice. W-N: well-nourished control. M-N: malnourished control. Re7d: renourished control. IEL: intraepithelial leukocytes.

1Significant differences with W-N control. 2Significant differences with M-N control. 3Significant differences with Re7d control. *P<0.05, **P<0.01 (ANOVA and Student’s Test).

●: IEL; ■: Goblet cells; ▼: Mast cells.
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function, IgA coated bacteria expressing analogous mannose rich glycans are prevented from adhering to epithelial cells without the need for specific antibody activity (Wold et al., 1990). We showed that the increase in the number of IgA+ cells in the lamina propria of intestine by L. casei was dose-dependent. This might provide a better intestinal barrier by immune exclusion against different enteropathogens. In addition, the concept of a common or integrated mucosal immune system (Mestecky et al., 1994) implies that oral immune stimulation can induce immunity in mucosal sites remote from the gut, such as other mucosal sites, e.g. respiratory and urogenital tissues, and mammary and salivary glands. On the other hand, IgA can trigger monocytes to show increased activity such as TNF-α secretion (Devière et al., 1991) and it can also cause eosinophil degranulation, which induces enhancement of the inflammatory response and the subsequent tissue damage. In our model, which has impaired mucosa by malnutrition, the S-IgA population must be carefully controlled because the L. casei feeding might become harmful, leading to tissue damage and promoting gut

![FIGURE 3. Photographs of hematoxilin-eosin staining of histological samples of small intestine. a: well-nourished control (X 40). b: malnourished control (X 40). c: renourished control (X 40). d: renourished mice treated with concentration of 10^7 cfu/day/mouse during 5 consecutive days of feeding (X 40).](image-url)
barrier compromise. The locally produced IgA is probably crucial for the immunological homeostasis within the lamina propria because this isotype lacks potent effector functions such as classical complement activation (Brandtzaeg et al., 1993). The increase of the number of IgG producing cells observed with some doses of *L. casei* feeding would indicate an inflammatory response because the mucosal IgG and IgM antibodies activate complement and may thereby impair the surface epithelium and enhancement of intestinal permeability.

Within the normal intestinal epithelium there is a large population of leucocytes, which are mostly accounted by lymphocytes (intraepithelial lymphocytes, IELs). It has been proposed the IEL have a number of different roles including: surveillance of the intestinal epithelial layer for the detection of microbial pathogens; removal of damaged or transformed epithelial cells; maintenance of epithelial integrity via secretion of trophic factors important for epithelial cell growth and differentiation; and the regulation of local cell mediated or humoral immune responses. (Aranda et al., 1999).

**FIGURE 4.** Transmission electron microphotographs of epithelial cell from small intestine. a: well-nourished control (X 4,900). b: malnourished control (X 12,800). c: renourished control (X 12,800). d: renourished mice treated with concentration of 10^7 cfu/day/mouse during consecutive 5 days of feeding (X 12,800).
The retention of T cells within the intra-epithelial space is very efficient and stable under normal conditions (Famularo et al., 1997). Several studies suggest that adhesion molecules and chemokine receptors that are selectively expressed on IELs are important for interactions with epithelial cells, such as the binding of integrin α E β 7 to E-cadherin, a ligand expressed on all epithelial cells. Finally these interactions of adhesion molecules on IEL with ligands on epithelial cells may modulate the functional response of either cell type (Agace et al., 2000). The decrease observed in the number of IEL after \textit{L. casei} feeding could be due to that it induces recovering of intestinal epithelium and would be more selective for retaining different immune cells. So, the administration of \textit{L. casei} might have more influence on epithelial cells than immune cells, and likewise it could avoid a powerful immune activation of IEL that can induce villus atrophy. By optic and transmission electron microscopy we showed that the supplementation of diet induces recovery of epithelial cells and mucosa, and we think it could influence the functionly of these cells.

It has been recently shown that T cells and mast cells share common homing/adhesion receptors, which suggests that they use similar migration pathways. Furthermore, these cells colocalize in the gastrointestinal mucosa, and it is possible that they act in concert to achieve an immune response (Smith and Weis, 1996). Mast cells contain a large number of bioactive mediators such as histamine and proteases and leukotrienes. However, recent work shows that mast cells can also play a critical role in innate immunity to bacterial infection and that mast cells and basophils can be activated by viral proteins (Wedemeyer et al., 2000). We did not observe significant variations when we studied the number of mast cells in the lamina propria after supplementation of the diet. It is important to inhibit the inappropriate activation of these cells because mucosal mast cells, together with eosinophils, T cells and other effector cells, constitute a network which can orchestrate protective immunity or harmful inflammation.

Goblet cells, present in both small and large intestine, release mucus granules from the apical cytoplasm after an inflammatory stimulus. Mucous secretion is triggered by direct stimulation by immune complexes and chemical agents and indirect stimulation by mediators released by histamine and lymphokines (Snyder and Walker, 1994). Therefore, the increase of the number of goblet cells observed in some doses of \textit{L. casei} administration could indicate an inflammatory response.

In conclusion this study showed that impaired gut barrier and mucosal immune function by malnutrition can be reversed by \textit{L. casei} used as an oral adjuvant of renutrition diet. The clinical significance of these findings will be important, in particular whether the chosen dose, as well as improving mucosal immunity, may also induce protection against enteropathogens.

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